Amendments to the Specification:

Please replace the paragraph beginning at page 113, line 4, as follows:

For the construction of Baculovirus expression vectors, the full-length human LPAAT-β cDNA was amplified by PCR from the DNA template pCE9.LPAAT-β (West *et al.*, *DNA Cell Biol. 16:*691-701 (1997)) using the primers 5'-TGATATCCGA AGAAGATCTT ATGGAGCTGT GGCCGTCTC-3' (olpb1F; SEQ ID NO:1) and 5'-CAGGCTCTAG ACTACTGGGC CGGCTGCAC-3' (olpb1R; SEQ ID NO:2). The ~870 bp fragment generated was reamplified by PCR using the primers 5' CCTACGTCG ACATGGAACA AAAATTGATA TCCGAAGAAG ATC-3' (olpb2F; SEQ ID NO:3) and 5'-CAGGCTCTAG ACTACTGGGC CGGCTGCAC-3' (olpb1R; SEQ ID NO:2). The ~890 bp fragment generated was then cleaved with Sal I and Xba I for insertion into pFastBac™ HTc vector (Life Technologies, Gaithersberg, MD) between the Sal I and Xba I sites for the generation of the plasmid pFB.LPAAT-β. This plasmid was then transformed into *E. coli* DH10Bac™ (Life Technologies, Gaithersberg, MD) for the generation of recombinant Bacmid DNA for transfection in HighFive (Invitrogen, San Diego, CA) or SF9 insect cells for the production of recombinant Baculovirus stocks using the protocol described in the Bac-to-Bac® Baculovirus Expression System (Life Technologies, Gaithersberg, MD), a eukaryotic expression system for generating recombinant baculovirus through site-specific transposition in *E. coli.* Viral stocks harvested from the transfected cells can then be used to infect fresh insect cells for the subsequence expression of LPAAT-β fusion protein with a poly-histidine tag and a myc-epitope near its N-terminus. The membrane fraction from these Sf9 cells would be the source of LPAAT enzyme.

Please insert the enclosed paper entitled "Sequence Listing" immediately after the section of the specification entitled "Abstract of the Disclosure" on page 143.